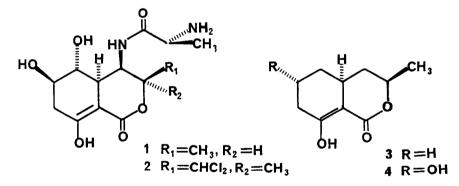
A SYNTHETIC APPROACH TO ACTINOBOLIN. TOTAL SYNTHESIS OF (±)-RAMULOSIN

Robert Cordova and Barry B. Snider^{*1} Department of Chemistry, Brandeis University Waltham, Massachusetts 02254

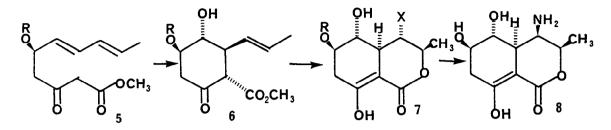
<u>Abstract</u>: A biomimetic type synthesis of (\pm) -ramulosin from the acyclic precursor <u>9</u> is described. Mercury mediated oxidative cyclization and bromolactonization are shown to be useful for the construction of the actinobolin skeleton.

The antibiotic actinobolin (1) was isolated from a <u>Streptomyces</u> culture 25 years ago.² Its structure was established in 1967 by Munk, Haskell and coworkers³ and confirmed in 1975 by x-ray crystallography.⁴ The closely related antibiotic bactobolin (2) was recently isolated from a <u>Pseudomonas</u> species.⁵ The potent antitumor activity of bactobolin and the novel structures of these antibiotics make them important synthetic targets.⁶ Two much simpler but biogenetically related compounds, ramulosin (3)^{7,8} and hydroxyramulosin (4)⁹ have been isolated from the fungus <u>Pestalotra ramulosa</u>.

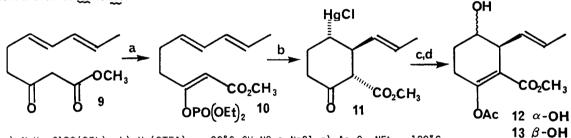


Our approach to the synthesis of actinobolin is based on an analysis of the likely biogenetic pathway. An acyclic pentaketide precursor 5 will be cyclized to the cyclohexanonecarboxylate 6. Halolactonization of 6 will give 7. The amine will then be introduced to give desalanylactinobolin 8 which can be easily converted to actinobolin (1).

Our initial studies, making use of the model compound 9, were designed to demonstrate the feasibility of the oxidative cyclization of 5 to give 6. The obvious approach, formation of the 6,7-epoxide of 9 and cyclization was not examined since ring closure of similar systems occurs on oxygen rather than carbon.¹⁰ Mercuric trifluoroacetate cyclization appears to suffer from similar problems,¹¹ although these can be solved by the use of the enol phosphate ester.¹² Methyl acetoacetate was converted to the dianion (NaH, BuLi)¹³ and treated with <u>E,E</u>-2,4-hexadienyl bromide¹⁴ to give <u>9</u>¹⁵ in 79% yield.



Treatment of 9 with NaH and diethyl chlorophosphate in ether¹² gave a 94% yield of 10. Treatment of 10 with 1.2 equiv. of mercuric trifluoroacetate in nitromethane at -20°C for 1 hour followed by workup with brine gave a 68% yield of the crystalline chloromercury compound 11, mp 183°C decomp.¹⁶ Oxidative demercuration of 11 was carried out by a twostep procedure. Protection of the ketone as the enol acetate (Ac₂0, Et₃N, 120°C, 30 min) followed by treatment with sodium borohydride in oxygen saturated DMF ^{12,17} gave a readily separable 3:1 mixture of 12 and 13 in 40% yield. These results indicate that mercury mediated cyclization followed by oxidative demercuration is a viable route for the conversion of 5 to 6.



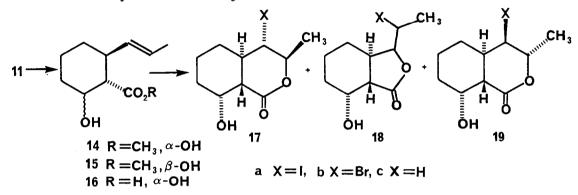
a) NaH, C1PO(OEt)₂ b) Hg(OTFA)₂, -20°C, CH₃NO₂; NaCl c) Ac₂O, NEt₃, 120°C d) NaBH₄, O₂, DMF

The halolactonization was explored with simpler models lacking the hydroxyl group. Reductive demercuration of 11 with NaBH₄ in EtOH gave a 50% yield of 14¹⁶ and a 38% yield of 15, mp 39-40°C.¹⁶ Hydrolysis of the ester of 14 gave 16, mp 86.5-87.5°C¹⁶ in 96% yield. Iodolactonization can give either the γ -lactone or the δ -lactone. In simple systems, the more stable γ -lactone is the major or exclusive product.¹⁸ Iodolactonization of 16 with I₂ and KI in aqueous NaHCO₃ solution gave a 91% yield of a 1:4:2 mixture of 17a, 18a (stereochemistry undetermined), and 19a. Iodolactonization with I₂ in ether-THF/aqueous NaHCO₃ solution¹⁹ gave an 86% yield of a 3:2 mixture of 18a and 19a. Iodolactonization thus leads primarily to the γ -lactone and in addition produces at best a mixture of diastereomeric δ -lactones favoring the undesired isomer.

We were pleased to find that bromolactonization of <u>16</u> with Br_2 in MeOH containing NaHCO₃ at -78°C²⁰ gave a readily separable 3.5:1 mixture of <u>17b</u> and <u>18b</u> in 88% yield. The stereochemistry of <u>17b</u>, mp 136-137°C, was unambiguously determined by analysis of the 500 MHz NMR spectrum.¹⁶ As a further proof of structure, <u>17b</u> was reduced with tri-<u>n</u>-butyltin

hydride in THF for 26 hr at $25^{\circ}C^{20}$ to give, after chromatography, a 76% yield of 17c, mp 116-117°C. Oxidation of 17c with $CrO_3 \cdot Pyr_2$ (freshly prepared <u>in situ</u>) in CH_2Cl_2 for 1 hr at $25^{\circ}C^{8,21}$ gave a 78% yield of (±)-ramulosin (3), mp 115-116°C, which is identical to natural (+)-ramulosin by 500 MHz ¹H NMR, IR and UV spectral, and TLC comparison.²²

These studies demonstrate an effective approach for forming the skeleton of actinobolin via sequential cyclization initiated by electrophilic attack on alkenes. They have led to the first synthesis of (\pm) -ramulosin in 7 steps from methyl actoacetate in 11% overall yield. The application of this approach to the synthesis of more highly oxidized members of this family is under investigation.



<u>Acknowledgement</u>: We thank the National Institutes of Health and the National Science Foundation (fellowship to R.C.) for financial support. The 500 MHz NMR spectrometer was purchased with funds provided by NIH grant GM 20168.

REFERENCES

- Fellow of the Alfred P. Sloan foundation 1979-1983. Dreyfus Teacher-Scholar 1982-1987.
- 2. T. H. Haskell and Q. R. Bartz, Antibiot. Ann., 1958-1959, 505 (1959).
- (a) F. J. Antosz, D. B. Nelson, D. L. Herald, Jr., and M. E. Munk, <u>J. Am. Chem. Soc.</u>, <u>92</u>, 4933 (1970).
 (b) M. E. Munk, D. B. Nelson, F. J. Antosz, D. L. Herald, Jr., and T. H. Haskell, <u>J. Am. Chem. Soc.</u>, <u>90</u>, 1087 (1968).
 (c) D. B. Nelson and M. E. Munk, <u>J. Org. Chem.</u>, <u>35</u>, 3832 (1970).
 (d) M. E. Munk, C. S. Sodano, R. I. McLean and T. H. Haskell, J. Am. Chem. Soc., <u>89</u>, 4158 (1967).
- J. B. Wetherington and J. W. Moncrief, <u>Acta Cryst.</u>, <u>B31</u>, 501 (1975) and references cited therein.
- (a) S. Kondo, Y. Horiuchi, M. Hamada, T. Takeuchi, and H. Umezawa, <u>J. Antibiotics</u>, <u>32</u>, 1069 (1979).
 (b) N. Ezaki, S. Miyadoh, T. Hisamatsu, T. Kasai and Y. Yamada, <u>J. Antibiotics</u>, <u>33</u>, 213 (1980).
 (c) I. Ueda, T. Munakata and J. Sakai, <u>Acta Cryst.</u>, <u>B36</u>, 3128 (1980).
- A total synthesis of (+)-actinobolin has been reported: M. Yoshioka, H. Nakai and M. Ohno, J. Am. Chem. Soc, 106, 1133 (1984).

- 7. F. H. Stodola, C. Cabot and C. R. Benjamin, <u>Biochem. J.</u>, <u>93</u>, 92 (1964).
- For a synthesis of (±)-epiramulosin see: J. A. Findlay, J. M. Matsoukas and J. Krepinsky, <u>Can. J. Chem.</u>, <u>54</u>, 3419 (1976).
- S. W. Tanenbaum, S. C. Agarwal, T. Williams and R. G. Pitcher, <u>Tetrahedron Lett.</u>, 2377 (1970).
- 10. V. N. Yandovskii and B. A. Ershov, <u>Russ. Chem. Rev</u>., (Engl. Transl.) <u>41</u>, 403 (1972).
- 11. T. R. Hoye, A. J. Caruso and M. J. Kurth, <u>J. Org. Chem.</u>, <u>46</u>, 3550 (1981).
- 12. E. J. Corey, M. A. Tius and J. Das, J. Am. Chem. Soc., 102, 1742 (1980).
- 13. S. N. Huckin and L. Weiler, <u>J. Am. Chem. Soc., 96</u>, 1082 (1974).
- 14. M. Jacobson, J. Am. Chem. Soc., 77, 2461 (1955).
- 15. D. Tunemato, N. Araki, K. Kondo, <u>Tetrahedron Lett.</u>, 109 (1977).
- 16. 500 MHz NMR $(CDCl_3) \delta 11 3.22 (d, J = 11.5 Hz, CHCO_2)$, 3.10 (ddd, J = 12, 11.5, 8.5 Hz, HCC=C), 2.81 (ddd, J = 12, 12, 3 Hz, CHHg); 14 4.14 (br dd, J = 2.5, 1.8 Hz, CHOH) 2.60 (dddd, J = 11.5, 11, 8.5, 3.5 Hz, HCC=C), 2.28 $(dd, J = 11, 1.8 Hz, CHCO_2)$; 15 3.78 (ddd, J = 11, 9.8, 4.3 Hz, CHOH), 2.18 (dddd, J= 12, 11, 6.6, 3.5 Hz, HCC=C), 2.11 $(dd, J = 11, 9.8 Hz, CHCO_2)$; 17b 4.52 (m, 1, CHOH), 4.51 (dq, J = 10.5, 6.2 Hz, CHOC), 3.62 (dd, J = 10.5, 10.5 Hz, CHBr), 2.29 (dddd, J = 12.5, 12.0, 10.5, 3.3 Hz, CHCBr); 2.09 (dd, J = 12.5, 1.8 Hz, CHCO).
- 17. C. L. Hill and G. M. Whitesides, J. Am. Chem. Soc., <u>96</u>, 870 (1974).
- For reviews see: (a) M. D. Dowle and D. I. Davies, <u>Chem. Soc. Rev.</u>, 171 (1979). (b)
 V. I. Staninets and E. A. Shilov, <u>Russ. Chem. Rev.</u>, (Engl. Transl.), <u>40</u>, 272 (1971).
- A. R. Chamberlin, M. Dezube, P. Dussault, and M. C. McMills, <u>J. Am. Chem. Soc.</u>, <u>105</u>, 5819 (1983).
- A. Sato, A. Ogiso, H. Noguchi, S. Mitsui, I. Kaneko and Y. Shimada, <u>Chem. Pharm.</u> Bull., 28, 1509 (1980).
- 21. A. B. Smith, III, P. A. Levenberg, Synthesis, 567 (1981).
- 22. We thank Dr. C. W. Hesseltine, Northern Regional Research Laboratory, USDA, Peoria, Ill. for a sample of natural ramulosin and Prof. J. A. Findlay for spectral data for ramulosin and epiramulosin.

(Received in USA 12 April 1984)